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## Note

### Separation of some androgens and estrogens by thin-layer chromatography

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During the synthesis of some new androgens ( $3\beta,17\beta$ -dihydroxy-16-oximino-5-androstene;  $3\beta,17\beta$ -dihydroxy-16,17-seco-5-androsteno-16-nitrile;  $3\beta,17\beta$ -dihydroxy-17 $\alpha$ -picolyl-5-androstene;  $3\beta$ -acetoxy-17 $\beta$ -hydroxy-17 $\alpha$ -picolyl-5-androstene;  $3\beta$ -hydroxy-17-picolinyliden-5-androstene;  $3\beta$ -acetoxy-17-picolinyliden-5-androstene;  $3\beta,17\beta$ -dihydroxy-16-oximino-17 $\alpha$ -picolyl-5-androstene)<sup>1</sup> and estrogens (17 $\beta$ -hydroxy-16-oximinoestrone 3-methyl ether; 16,17-seco-17-oxoestrone-16-nitrile 3-methyl ether)<sup>2</sup>, mixtures of particular derivatives were obtained. For the control of the synthesis, it was necessary to identify these derivatives.

Identification was carried out by thin-layer chromatography (TLC) on silica gel layers. Various solvent systems for steroid separations have been described<sup>3-13</sup>, but none of them was found to be completely adequate for the separation of the particular mixtures in which we were interested.

This paper describes some modified developing systems for the TLC of the steroids examined.

## EXPERIMENTAL

For the preparation of the thin layers, silica gel G (Merck, Darmstadt, G.F.R.) was used. Silica gel (30 g) was suspended in 75 ml of distilled water and the suspension was coated on to glass plates (20 × 20 cm) with Desaga equipment. The thin layers were dried in air at room temperature and activated for 1 h at 110°.

The mixture of eleven androgens (Table I) and the mixture of five estrogens (Table II) were examined. Solutions of each steroid in chloroform (0.5%) were prepared, and 0.5  $\mu$ l of each steroid solution and the mixtures were applied to the chromatoplate with a micropipette.

The chromatograms were developed at room temperature in a glass chamber containing 50 ml of solvent mixture without previous saturation, by one- and two-dimensional ascending chromatography. The following solvent systems were used for the separation of androgens:

- (A) benzene-acetone (7:1) in the first dimension;
- (B) benzene-ethyl acetate (2:1) in the second dimension; and for the estrogens:
- (C) benzene-ethyl acetate (7:1);
- (D) benzene-acetone (8:1);



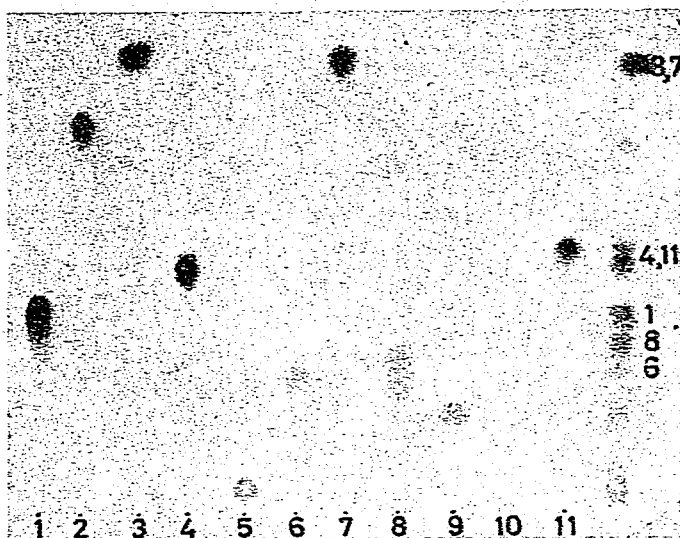


Fig. 2. Chromatogram of androgens 1-11 (Table I) in solvent system B.

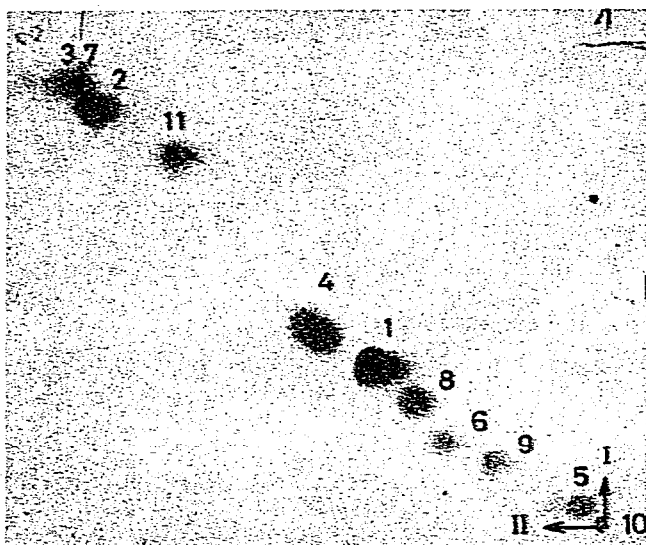


Fig. 3. Two-dimensional chromatogram of androgens 1-11 (Table I). Run I, solvent system A; run II, solvent system B.

- (E) chloroform-ethyl acetate (4:1);
- (F) *n*-hexane-acetone (2:1);
- (G) carbon tetrachloride-ethyl acetate (2:1).

The developed and dried chromatograms were developed by spraying with 50% sulphuric acid in methanol and heating in an oven for 10-15 min at 100-110°.

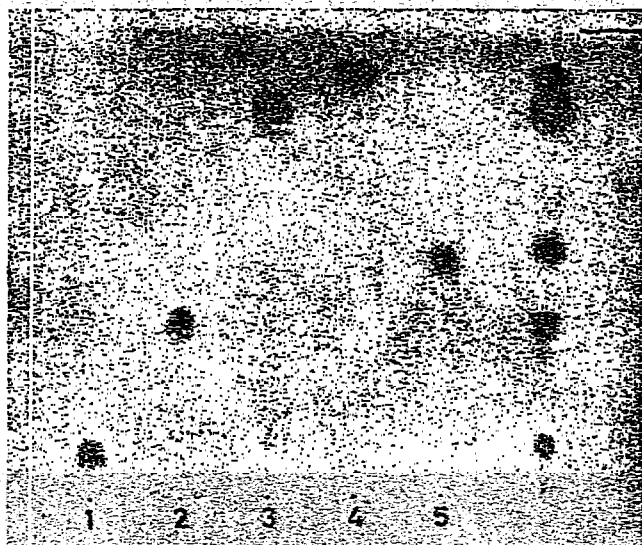


Fig. 4. Chromatogram of estrogens 1-5 (Table II) in solvent system C.

## RESULTS

Both solvent systems A and B clearly resolved nine androgens by one-dimensional chromatography (Figs. 1 and 2). The  $R_F \times 100$  values for each steroid are recorded in Table I. The same solvents separated ten androgens by two-dimensional chromatography (Fig. 3), but no separation of  $3\beta$ -acetoxy-17-picolinylyden-5-androstene and  $3\beta$ -acetoxy-5-androsten-17-one was obtained.

All estrogens were clearly resolved by one-dimensional chromatography with the solvents used (Table II). The best separation was obtained with solvent system C (Fig. 4).

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